

### **AMENDMENTS TO THE SPECIFICATION**

Please replace the paragraph on page 34, lines 14-20, with the following paragraph:

Ordinarily, an aqueous aerosol is made by formulating an aqueous solution or suspension of the agent together with conventional pharmaceutically-acceptable carriers and stabilizers. The carriers and stabilizers vary with the requirements of the particular compound, but typically include nonionic surfactants (~~Tweens~~ TWEEN®, ~~Pluronics~~ PLURONIC®, or polyethylene glycol), innocuous proteins like serum albumin, sorbitan esters, oleic acid, lecithin, amino acids such as glycine, buffers, salts, sugars or sugar alcohols. Aerosols generally are prepared from isotonic solutions.

Please replace the paragraph on page 43, lines 13-33, with the following paragraph:

For HIV tetramer formation, biotinylated DR/peptide complexes were incubated with R-phycoerythrin-labeled streptavidin (Molecular Probes, Eugene, OR) for at least one (1) hour on ice, at a 4:1 molar ratio of DR to streptavidin and a final DR concentration of 0.2 mg/ml. For analysis of blood lymphocytes from patients infected with HIV, CD4<sup>+</sup> T cells were enriched using the CD4<sup>+</sup> RosetteSep™ reagent (StemCell Technologies, Vancouver, BC, Canada). This process results in binding of unwanted cell populations to red blood cells, which pellet during Ficoll FICOLL® density gradient centrifugation. Four-color FACS analyses (Using a FACSCalibur™ machine (Becton Dickinson) and CellQuest™ software) was performed with the PE-labeled tetramer and antibodies to CD3 (FITC-labeled), CD4 (PerCP-labeled), and CD25 (APC-labeled) (all acquired from BD Pharmingen, San Diego, CA), which allowed gating on the CD3<sup>+</sup> population and analysis of CD4<sup>+</sup> or CD25<sup>+</sup> versus tetramer staining. Isolated CD4<sup>+</sup> T cells were stained with tetramers (typically at 10 µg/ml) in RPMI, 10% human serum for one hour at 37°C in the presence of labeled antibodies, washed twice and fixed in 1% formaldehyde/PBS. For staining of T cell lines, monocytes were depleted by Ficoll FICOLL® density gradient centrifugation with a CD4<sup>+</sup> enrichment cocktail (StemCell Technologies) and staining was performed with 100 µl of PBS with 2% fetal calf serum in the presence of tetramers and labeled antibodies. Lymphocytes were gated based on forward and side scatter and CD3 expression. Multiple control tetramers with irrelevant peptides were used in most experiments to demonstrate specificity of binding by the relevant tetramers (see Table 2).